Reviews

Diastereoselective construction of small building blocks via [2+2] cycloadditions involving ketenes: A direct incorporation of α -, β -, and γ -amino acids into peptides*

Claudio Palomo,* Jesus M. Aizpurua and Inaki Ganboa

Departmet of Organic Chemistry, Basque Country University, Faculty of Chemistry, **
Apdo, 1072. 20080 San Sebastian, Spain

Data concerning diastereoselective construction of peptides via β -lactams obtained from [2+2] cycloaddition reactions with participation of ketenes are summarized.

Key words: α -amino acid N-carboxy anhydrides, β -amino- α -hydroxy acids, α,β -diamino acids, macrocyclic peptides, 1,3-polyols and amino polyols, pyrrolizidine alkaloids, β,γ -dihydroxy acids, γ -amino- β -hydroxy acids.

1. Introduction

There are a number of reasons for the current interest in the synthesis of unusual amino acids, particularly the need to prepare peptidomimetics, synthetic enzymes, and new drugs (see reviews²⁻⁷). As a result, a number of suitable methods for the synthesis of both α - and β -amino acids have been developed over the last few years. Several comprehensive reviews concerning syntheses of both α -amino acids⁸⁻¹² and β -amino acids have been published. 13,14 However, most of the investigations on this subject deal with the synthesis of the non-proteinogenic amino acids in their free forms, rather than with the generation of simultaneously N-protected and COOH-activated species ready for subsequent peptide coupling. Towards this goal Heimgartner has devel-

Carboxyl group activation has been accomplished, on one hand, through the formation of α,α -dialkyl oxazolones and their coupling with α -amino acid esters (Scheme 2)¹⁶ and, on the other hand, by the *in situ* generation of amino ketenes as reactive species. In this context, the photolysis of chromium α -aminocarbene complexes and their subsequent coupling with α -amino acid esters¹⁷ (Scheme 3) as well as the chain extension of α -amino acids *via* the Arndt-Eistert reaction and trapping of the resulting β -amino diazoketenes with α -amino acid esters and dipeptides^{18,19} (Scheme 4) were developed. A single-step synthesis of racemic di- and tripeptides from unnatural β -hydroxy- and β -mercapto α -amino acids by the Ugi reaction has also been recently described.²⁰

During the course of our investigations on β -lactams we discovered that Baeyer—Villiger rearrangement (Scheme 5) of racemic α -keto- β -lactams (2) readily

oped the azirine-oxazolone method (Scheme 1), putting into practice the idea of amino group activation and providing a way for the incorporation of α,α -disubstituted α -amino acids into peptides. 15

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Scheme 2

Scheme 3

Reagents and conditions: i, hv, 0 °C, THF, CO (4-5 atm).

obtainable from α -hydroxy- β -lactams (1) takes place regioselectively to give α -amino acid N-carboxy anhydrides (NCAs) (3). The well recognized importance of this particular class of mixed anhydrides in α -amino acid chemistry 22,23 led us to develop this approach into a general method for the synthesis of homochiral NCAs that would proceed in a single one-pot operation without the need to isolate the intermediate α -keto- β -lactams. We found that the stable nitroxide free radical, 2,2,6,6-tetramethylpiperidinyl-1-oxyl (TEMPO), in combination with a solution of commercial bleach fulfills this criterion and provides NCAs from α -hydroxy- β -lactams in yields higher than 95%. In this context,

Scheme 4

Reagents and conditions: i. 1) NEt₃, ClCO₂Et; 2) H₂CN₂;

ii.
$$H_2N$$
 CO_2Bn , 10% AgO₂CPh, NEt₃.

much of our work has been centered on [2+2] cycload-dition reactions of α -hydroxyketene equivalents with imines to obtain the required α -hydroxy- β -lactams in a convergent fashion (a general review on β -lactams (Ref. 25) and a comprehensive review concerning cycloaddition of ketenes to imines (Ref. 26) should be mentioned), although ester enolate-imine condensation could also be employed for the same purpose. ^{27,28}

Besides being useful as valuable precursors of NCAs, α -hydroxy- β -lactams have also been found to be excel-

Reagents, conditions, and yields: *i.* DMSO, P_2O_5 , 16 h, ~20 °C, 89-90%; *ii.* m-ClC₆H₄CO₃H (MCPBA), CH₂Cl₂, -40 °C, 1 h, 80-90%; *iii.* NaOCl, TEMPO, > 95%.

lent building blocks of α -hydroxy- β -amino acid derivatives by exploiting nucleophilic reactions at the β -lactam carbonyl group (see comprehensive reviews 29-32). When we initiated our investigations on this subject, little was known about β -lactam ring opening by carbon nucleophiles³³⁻³⁹ and even less about coupling reactions with α -amino acid esters.^{40,41} Therefore, we studied both aspects with the aim of establishing new reaction methodology for the incorporation of β -amino acid derivatives into peptides.

This account summarizes our studies on α -, β -, and γ -amino acid peptides and related systems derived from β -lactams as readily available synthetic building blocks.

2. Synthesis of \alpha-amino acid N-carboxy anhydrides

 α -Amino acid N-carboxy anhydrides, NCAs, or Leuchs anhydrides, offer both amino group protection and carboxylate activation simultaneously. As a consequence, they have found wide application in peptide synthesis and numerous procedures have been reported for their preparation, all of them involving reactions between an α -amino acid and dehydrating agents. $^{42-51}$ Therefore, we considered that a conceptually new approach to peptide segments would be facilitated if NCAs could be obtained from non- α -amino acid precursors. To this end, we have developed the first tactically new approach to NCAs, proceeding by an unprecedented ring expansion of α -hydroxy- β -lactams (this topic is treated in a review, 52 another method for ring expansion of α -hydroxy- β -lactams is described in Ref. 53).

2.1. α,β -Diamino acid N-carboxy anhydrides

2,3-Diamino carboxylic acids are uncommon naturally occurring amino acids that have aroused consider-

able interest (see Refs. 54—62). This particular class of amino acids, when incorporated into peptides with either the β - or the α -amino group as part of the backbone, might provoke changes in polarity, additional hydrogenbonding interactions, and may make the peptide basic. α,β -Diamino acids can also serve as substitutes for α -hydroxy- β -amino acid-derived peptides to probe their structural specificity and topology. ⁶³ Interest in α,β -diamino acids also stems from their occurrence in cyclic peptides and biologically active substances, e.g. capreomycin, ⁶⁴ pyrimidoblamic acid, ^{65,66} auisqualic acid, ^{67,68} willardiine, ⁶⁹ mimosine. ⁷⁰ β -Substituted 2,3-diamino acids are also useful intermediates for the synthesis of imidazolines, which have been employed as amide bond replacements in the design of peptidomimetics. ^{71–74}

The approach to α,β -diamino acid N- α -carboxy anhydrides (Scheme 6) takes advantage of the highly diastereoselective cycloaddition of ketenes to imines derived from N-Boc- α -amino aldehydes (Boc = ButOCO) developed in our laboratory. 75 The method 24 involves a one-pot double oxidation sequence of α-hydroxy-β-lactams (5) using TEMPO in combination with fresh commercial bleach whose pH = 12.7 was adjusted to neutrality in order to avoid possible epimerization of the resulting NCA (6). Reactions can be conducted at 0 °C and are almost complete in a few minutes using a twofold excess of 1 M NaOCl and a catalytic amount of TEMPO. Without the catalyst, the starting α-hydroxy-β-lactams are recovered unchanged after 24 h of reaction at room temperature. As shown in the scheme, the method allows the formation of (R)- α -amino acids (7) from (S)- α -amino aldehydes (imi-

Scheme 6

Reagents and conditions: i. 1) BnOCH₂COCl, NEt₃, CH₂Cl₂, 75–85%; 2) H₂, Pd/C, EtOH, 90–95%; ii. 1 N NaOCl, TEMPO, NaHCO₃, KH₂PO₄-K₂HPO₄, KBr (pH 6.9), CH₂Cl₂, -20 °C, 10–15 min, 90–98%; iii. 1) MeOH, boiling, 72–90%, 2) H₂, 10% Pd/C, EtOH.

nes 4) and vice versa with completely predictable stereochemical control and optical purity. Furthermore, both amino moieties are differentially protected and thus incorporation of these amino acids into peptide chains either at the α - or β -positions becomes possible via this procedure.

The use of the enantiomers of imines 4 makes it possible to obtain the corresponding enantiomeric azathreonines and peptides thereof. This latter aspect is illustrated (Scheme 7) by the synthesis of aminodeoxybestatin 11, an inhibitor of aminopeptidase-M equipotent to the known bestatin. 63 For example, imine 8, on treatment with benzyloxyacetyl chloride and triethylamine followed by hydrogenolysis of the resulting 3-benzyloxy-β-lactam, led to hydroxylactam (9) in good yield. Treatment of 9 with TEMPO and NaOCl as above gave the NCA 10 in 97% yield, which on coupling with (S)-leucine methyl ester furnished the corresponding dipeptide product. Subsequent N-debenzylation of the latter led to 11 in good overall yield. For the synthesis of azathreonine-NCAs in a two-step procedure see Ref. 76.

Scheme 7

Reagents, conditions, and yields: i. 1) BnOCH₂COCl, NEt₃, 2) H₂, Pd/C, 90%; ii. 1 N NaOCl, TEMPO, 97%;

Along with these studies we also examined an alternative β -lactam approach to these α,β -diamino acid peptides, which will be discussed in section 3.2.

2.2. α -Amino- β -hydroxy acid N-carboxy anhydrides

The above reaction methodology has also been extended to threonine NCAs⁷⁷ as a potential novel route to peptide segments of macrocyclic antibiotics. For example (Scheme 8), treatment of imines (12a—c) with benzyl-

oxyketene and further debenzylation of the resulting cycloadducts led to the α -hydroxy- β -lactams (13a-c) in good yields. Subsequent oxidation of each compound 13 with P_2O_5 in DMSO gave the corresponding α -keto- β -lactams (14). We have also observed that reduction of these α -keto- β -lactams with sodium borohydride proceeds with complete stereoselectivity to give the starting α -hydroxy- β -lactams 13, thus proving the lack of epimerization during their oxidation. Finally, Baeyer—Villiger rearrangement of each α -keto- β -lactam 14 furnished the expected NCAs (15) in excellent yields although, in some instances, they were contaminated with m-chloroperbenzoic acid. Nonetheless, the direct one-pot transformation of α -hydroxy- β -lactams into the desired NCAs has been found to be the method of choice.

Scheme 8

Reagents, conditions, and yields: i. 1) BnOCH₂COCl (2 equviv.), NEt₃, 70–85%; 2) NH₄CO₂H, Pd/C, PriOH, 89–93%; ii. P₂O₅, DMSO, ~20 °C, 20 h, 85–95%; iii: MCPBA, CH₂Cl₂, -40 °C, 1 h, 90–95%; iv. NaOCl, TEMPO (cat.), NaHCO₃, KH₂PO₄, K₂HPO₄ (pH = 6.9), CH₂Cl₂, > 95%.

The optical purity of these NCAs was determined (Scheme 9) by their prior conversion into the respective

Scheme 9

Reagents and conditions: i. 1) MeOH, boiling, 1 h; 2) NH₄CO₂H, Pd/C; 3) (+)-MTPA—Cl, NEt₃.

a: R = Me; b: R = Bn; c: R = Pr

Reagents, conditions, and yields: i. 1) BnOCH₂COCl (2 equviv.), NEt₃, 70–85%; 2) NH₄CO₂H, Pd/C, PrⁱOH, 89–93%; ii. P₂O₅, DMSO, ~20 °C, 20 h, 80–92%; iii. MCPBA, CH₂Cl₂, 80–90%; iv. NaOCl, TEMPO (cat.), NaHCO₃, KH₂PO₄, K₂HPO₄ (pH = 6.9), CH₂Cl₂, > 95%.

O-protected threonines and subsequent acylation with Mosher acid chloride ((+)-MTPA-Cl) to form compounds 16.

Owing to the importance of α -methyl- β -alkylserines for the study and design af new bioactive targets, ^{78–80} the above approach has also been developed into a general method for the synthesis of α -branched α -amino- β -hydroxy acid N-carboxy anhydrides. ⁸¹ The key to the approach was the use of α -alkoxyketone-derived imines (17). These are readily available starting materials that

incorporate the required structural subunit of the desired amino acid and, at the same time, provide chirality to the corresponding NCA precursors (Scheme 10). When we started this work, no general method for the synthesis of homochiral β-lactams with quaternary stereogenic centers at C(4) was available. For this reason we explored the cycloaddition of benzyloxyketene with the aforementioned imines in the hope of obtaining the expected cycloadducts with high diastereoselectivity. Indeed, using standard cycloaddition conditions we got the corresponding \beta-lactams in good yields and, most notably, as single diastereomers. Subsequent removal of the benzyloxy protective group from the resulting cycloadducts led to the α -hydroxy- β -lactams (18). The absolute configuration assigned to the adducts was based initially on the assumption of a uniform reaction mechanism and then was confirmed by a single crystal X-ray analysis of 18c. Conversion of these adducts into the corresponding NCAs 20 was first carried out in a twostep procedure involving oxidation of each β-lactam 18 to the respective α-keto-β-lactam 19 and further Baeyer— Villiger rearrangement, but better yields were obtained by performing this oxidation sequence in a one-pot operation using TEMPO in combination with NaOCI. In these instances, the insertion of the oxygen atom also takes place regioselectively between both carbonyl groups.

After the preparation of α -branched α -amino- β -hydroxy acid NCAs, we explored their coupling reactions with several α -amino esters, but problems arose leading to yields below 10%. However, as Scheme 11 illustrates, the desired coupling reactions of α -amino esters with the above NCAs were achieved efficiently using potassium cyanide as additive. Under these conditions amino esters 21 and 22 coupled with 20a to give dipeptides 23 and 24 in 87 and 90% isolated yields. In a similar way α -amino acid esters 25 and 26 gave 28, 29

Reagents, conditions, and yields: i. 20a, KCN, DMF, 90%; ii. 20b, KCN, DMF, 90%; iii. 20c, KCN, DMF, 79-90%.

and 30 in good yields too. Under these conditions even the bulky Aib-methyl ester 27 coupled with 20c to give 31 in 79% yield.

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The potential of the β -lactam-NCA method is also demonstrated by the synthesis of aryl- and homoarylalanines. The key to the approach (Scheme 12) is the use of the N,O-protected β -formyl- β -lactam 34 as a common homochiral synthetic building block readily obtained from the β -lactam 33.

Scheme 12

Reagents, conditions, and yields: i. BnOCH₂COCl, NEt₃, 76%; ii. 1) 3 N HCl, MeOH, boiling, 75%; 2) NaIO₄, Me₂CO—H₂O, 24 h, ~20 °C, 95%.

Similarly, the enantiomer (35) of the imine 32 (Scheme 13), after N, O-dideprotection and subsequent oxidative cleavage, furnished the β -lactam 37. The latter

Scheme 13

Reagents, conditions, and yields: i. BnOCH₂COCI, NEt₃, 65-80%; ii. 1) TsOH, THF, H₂O, 90%; 2) NaIO₄, Me₂CO, H₂O, 95%.

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can also be produced in a similar way from Bose's cycloadduct 36 derived from the glyceraldehyde imine 35b.83 In both cases yields were generally high and the reactions were performed in gram quantities.

As Scheme 14 illustrates, the synthesis of homoarylalanine NCAs starts from aldehyde 37 by using the Wittig reaction followed by hydrogenolysis of the resulting olefinic intermediates under Pd/C. Subsequent oxidation and Baeyer-Villiger rearrangement of the α-ketoβ-lactam intermediates 39 afforded the desired NCAs 40 in good yields. Once again, considerable yield improvement (typically >95%) was observed by performing the direct transformation of 38 into 40 promoted by TEMPO. It is interesting to note that this approach involving β-formyl-β-lactam elongation also allows the synthesis of other structurally different NCAs depending on the preselected Wittig reagent. At the same time (Scheme 15), simple Grignard addition to the formyl- β -lactam 37 followed by deoxygenation of the resulting mixture of epimeric β-lactams 41 furnished 42 in good yields. One-pot double oxidation of these products gave NCAs 43 formally derived from arylalanines.

Scheme 14

a: $R = Ph, R^1 = H;$ **d:** $R = H, R^1 = H;$ **b:** $R = 4\text{-MeOC}_6H_4, R^1 = H$ **e:** $R = Me, R^1 = H$ **b:** $R = 4\text{-MeOC}_6H_4, R^1 = H$ **f:** $R = Me, R^1 = Me$

Reagents, conditions, and yields: i. 1) $Ph_3P = CHRR^1$, THF, ~ 20 °C, 2 h; 2) NH_4CO_2H , Pd/C, MeOH, 75-90%; ii. P_2O_5 , DMSO, ~ 20 °C, 20 ч, 70-90%; iii. MCPBA, CH_2Cl_2 , ~ 40 °C, 1 h, 90-95%; iv. NaOCl, TEMPO (cat.), $NaHCO_3$, KH_2PO_4 , K_2HPO_4 (pH=6.9).

In view of the increasing importance of α,α -disubstituted α -amino acids for the study and design of structurally defined peptides, peptidomimetics, and, in general, potent bioactive targets, $^{2-7}$ the above β -formyl- β -lactam elongation has also been extended to the synthesis of dipeptide segments containing this class of amino acids. The cycloaddition of benzyloxyketene to the

Reagents, conditions, and yields: i. 1) RMgBr, THF, -40 °C, 30 min, 72-88%; 2) AcCl, NEt₃, CH₂Cl₂, \sim 20 °C, 2 h, 95--100%; ii. NH₄CO₂H, Pd/C, PriOH, boiling, 1 h, total yield 71-90%; iii. NaOCl, TEMPO, pH = 6.9, >90%.

ketimine 44 leads to lactam 45 with perfect asymmetric induction at the newly created stereogenic centers. Subsequent deprotection of the acetonide group and further oxidative cleavage of the resulting diol gave formyl- β -lactam 46 in good overall yield (Scheme 16). With this material in hand, the synthesis of β , β -disubstituted α -hydroxy- β -lactams 47 was performed as above using the Wittig reaction, followed by hydrogenolysis of the benzyloxy protective group and simultaneous double bond reduction. As Scheme 17 shows, when these com-

Scheme 17

Reagents, conditions, and yields: i. BnOCH₂COCI, NEt₃, 80%; ii. 1) HClO₄, THF, H₂O; 2) NalO₄, Me₂CO, H₂O, 24 h, 90%.

Scheme 17

Reagents, conditions, and yields: *i.* 1) Ph₃P=CHRR¹, THF, ~20 °C, 2 h; 2) H₂, Pd/C, EtOH, ~20 °C, 14 h, 75%; *ii.* NaOCl, TEMPO (cat.), 95—97%; *iii.* 1) (S)-H₂NCR¹R²CO₂R³, KCN, DMF; 2) H₂, Pd/C, EtOH, 70—73%.

pounds were treated with NaOCl and a catalytic amount of TEMPO the corresponding NCAs 48 were formed in yields up to 97%. In accordance with the results outlined previously, NCAs 48a and 48b were resistant to ring opening by α -amino acid esters such as (S)-phenylalanine methyl ester and (S)-valine benzyl ester, but with the addition of potassium cyanide the coupling reaction proceeded cleanly to give, after hydrogenation over Pd on charcoal, dipeptides 49 and 50 in yields of 73 and 71%, respectively. Under these conditions, even the bulky benzyl α -aminobutyrate could be efficiently coupled with the NCA 48e to give the corresponding dipeptide derivative, which upon N, O-didebenzylation afforded dipeptide 51 in 70% yield.

3. B-Amino acids

In connection with the above studies on NCAs we also developed the use of β -lactams as synthetic equivalents of β -amino acids. It is well known that β -lactams undergo hydrolytic cleavage of the C-N amidic linkage to form β -amino acids under both acidic and basic conditions. ²⁹⁻³² However, the harsh reaction conditions often required to carry out this transformation can

cause partial or complete epimerization, not only at the α-position of the β-lactam ring, but also at other positions of the \(\beta\)-lactam product. This is particularly true for compounds of general structure 52 or 53, in which the exocyclic methyl ester group can, in its turn, be easily cleaved. In concert with these problems, little was known about the behaviour of \(\beta\)-lactams as acylating agents for peptide synthesis before we started our work on this subject. The only investigation on this topic was reported by Drey and his co-workers, who attempted the coupling reaction of a β-lactam framework with α-amino acid esters but with very little success. 40 At the same time, it is also well known that some simple monocyclic β-lactams, when carrying suitable electron-withdrawing groups at the N atom, possess antimicrobial activity and/or act as inhibitors of transpeptidases, \(\beta\)-lactamases. and elastases.87 Therefore, on this basis we reasoned that if the final result of the opening of the β-lactam ring by an enzyme results in its O-acylation, a parallel coupling reaction between monocyclic β-lactams and α-amino acid esters should be possible. If so, the net effect would be a conceptually new approach to peptide synthesis incorporating β-amino acid segments.

Our initial trials to probe this hypothesis were made on N-Boc-B-lactams and the finding was that the β-lactam ring could efficiently be opened by both oxygen and nitrogen nucleophiles in the presence of sodium azide or potassium cyanide as promoters of the reaction88 (see also Refs. 89, 90). Presumably an acyl azide or an acyl cyanide intermediate should be formed in such a reaction. Although no evidence has yet been found for this assumption, it remains a fact that β-lactam ring opening generally does not take place in the absence of these additives. On the other hand, since the overall process would imitate the alcoholysis of β-lactams promoted by class C β-lactamases, 91 the term enzymemimetic was adopted for these coupling reactions. This aspect will be illustrated in the following sections through some representative examples that add new perspectives to β-amino acid chemistry.

3.1. β -Amino α -hydroxy acids

The synthesis of β -amino- α -hydroxy acids to be incorporated into peptides has been well documented and still continues to be of current interest within the domain of new enzyme inhibitors. Proportion 12 Important members of this class of compounds are bestatin 54 and amastatin 55, two low molecular weight peptidic immunomodifiers 93,94 with antitumor and antimicrobial activity. Proposed 15 in the synthesis of the synthesis

The synthesis of these compounds requires the coupling of two structural units, the corresponding N-terminal β -amino- α -hydroxy acid and the C-terminal amino acid leucine or the C-terminal tripeptide Val-Val-Asp.

Our approach to (-)-bestatin 54 involves the cycloaddition of benzyloxyketene to the imine 56 (Scheme 18) according to the procedure of Terashima⁹⁶ and subsequent desilylation and deoxygenation of the resulting adduct 57 (PMP is p-methoxyphenyl) under

Scheme 18

Reagents, conditions, and yields: *i*. BnOCH₂COCl, NEt₃; *ii*. 1) Bu₄ⁿNF, CH₂Cl₂, $^{\circ}$ S%; 2) NaH, CS₂, THF, 0 $^{\circ}$ C, Mel, $^{\circ}$ 20 $^{\circ}$ C, 30 min, 100%; 3) Bu₃ⁿSnH, Et₃B, C₆H₆, $^{\circ}$ 20 $^{\circ}$ C, 60%.

modified Barton's conditions. The β -lactam 58, which is the cyclized form of the bestatin β -amino acid, is then N-dearylated under the action of cerium ammonium nitrate (CAN) and activated with the introduction of the Boc group to this position. At first (Scheme 19) the coupling reaction of the resulting N-Boc- β -lactam 59 with (S)-leucine benzyl ester was examined in methylene chloride at room temperature, but under these conditions a fourfold excess of the α -amino acid ester was required to give complete conversion into the de-

Reagents, conditions, and yields: i. 1) CAN, MeCN, H_2O , 90%; 2) $(Boc)_2O$, DMAP, MeCN, 73%; ii. $H_2N \frown CO_2Bn$, DMF, NaN₃ (1 equiv.), 20 h, ~20 °C, 88%; iii. 1) TFA, CH_2Cl_2 ; 2) H_2 , Pd/C, EtOH, 98%.

sired dipeptide product 60. However, when the reaction was performed in DMF and in the presence of NaN₃ the coupling proceeded by using equimolar amounts of both components⁹⁷ (see also Ref. 98).

This strategy was also employed for the synthesis of β -hydroxyaspartic acid derivatives. β -Hydroxyaspartic acid itself appears to play an important role in blood clotting proteins⁹⁹ and its derivatives have been found in macrocyclic antibiotics like lysobactin, 100-103 vide infra. The synthesis of the starting β -lactam (Scheme 20) was carried out from imines 61 and 62 via cycloaddition; in both cases one diastereomer was formed. The resulting

Scheme 20

Reagents, conditions, and yields: i. BnOCH₂COCl, NEt₃, 80-85%; ii. 1) HCl, MeOH, 100%; 2) NaIO₄, Me₂CO-H₂O, KMnO₄, -20 °C, 75%.

β-lactams 63 and 64 were then converted into the 4-carboxy derivative 65 in good overall yield. It should be mentioned that the use of chiral glyoxylates in this cycloaddition reaction afforded the corresponding 4-alkoxy-carbonyl β-lactams, albeit with poor diastereoselectivity.¹⁰⁴

The β -lactams 67a and 67b can be obtained by esterification of the acid 65, followed by N-dearylation of the esters 66a and 66b obtained, respectively (Scheme 21). Ring opening of the NH β -lactams 67 by means of chlorotrimethylsilane in methanol led to the corresponding β -benzyloxy aspartate 68b.

Scheme 21

BnO
$$\stackrel{\text{H}}{\longrightarrow}$$
 $\stackrel{\text{H}}{\longrightarrow}$ $\stackrel{\text{CO}_2R}{\longrightarrow}$ $\stackrel{\text{MeO}_2C}{\longrightarrow}$ $\stackrel{\text{N}}{\longrightarrow}$ $\stackrel{\text{N}}{\longrightarrow}$ $\stackrel{\text{CO}_2R}{\longrightarrow}$ $\stackrel{\text{N}}{\longrightarrow}$ $\stackrel{\text{R}}{\longrightarrow}$ $\stackrel{\text{R}}{\longrightarrow}$

Reagents, conditions, and yields: i. CAN, MeCN, H₂O, 70-80%; ii. 1) MeOH, ClSiMe₃; 2) (Boc)₂O, 70-75%.

At the same time, the introduction of the Boc group at the N-position of the β -lactam 67a led to the lactam 69, which then (Scheme 22) was coupled with (S)-phenylalanine and (S)-valine benzyl esters to give dipep-

tides 70 and 69, respectively. Both couplings proceeded efficiently in the presence of NaN₃ but, in its absence, only 50% conversion was observed in the reaction of lactam 69 with (S)-valine benzyl ester.⁹⁷

Scheme 22

3.2. α,β -Diamino acids

As pointed out in section 2.1, α,β -diamino acids are of particular interest in the development of new peptidomimetics and syntheses that allow their incorporation into peptide chains by either the α - or the β -amino function. These facts led us to prepare β -alkyl- α -amino- β -lactams with differently protected amino moieties and, in the light of the above results, to study their coupling with α -amino acid esters.

Although the cycloaddition reaction of imines with ketenes generated from acid chlorides and tertiary amines has provided a useful approach to diverse β-lactam compounds, it has not hitherto been viable with enolizable imines, in part because of their instability, and in part because of competitive deprotonations. 105 Efforts from this laboratory to solve this problem resulted in the development of imines 72, which, in spite of their remarkable stability, react with the Evans-Sjogren ketene XcCH=C=O (Xc = 2-oxo-4-phenyloxazolidin-3-yl), generated from the corresponding acid chloride and triethylamine, to form β-lactams 73 with excellent diastereomeric ratios and, most notably, with perfect asymmetric induction at the C(3) atom. 106 This method (Scheme 23) is applicable for the construction of homochiral \(\beta \)-lactams with linear as well as branched chains at the B-C atom. Removal of the chiral auxilliary followed by protection of the amino group led to the Cbz-derivatives 74, which by treatment with CAN in acetonitrile-water gave N-formyl-β-lactams 75. The latter were deformylated to the desired N-H β-lactams 76 under slightly basic conditions.

It was also found that an alternative removal of the N-bis(trimethylsilyl)methyl substituent with CAN in methanol directly afforded the corresponding NH β -lactams. However, the subsequent cleavage of the

Scheme 23

Reagents, conditions, and yields: i. XcCH₂COCl, NEt₃, CHCl₃, boiling, 20 h, 55–82%; ii. 1) Li, NH₃, Bu¹OH, THF, >94%; 2) CbzCl, DMAP, 70–75%; iii. (NH₄)₂Ce(NO₃)₆, MeCN, H₂O, 80%; iv. NaHCO₃, Na₂CO₃, H₂O, Me₂CO, 70–80%.

f: R1 = H. R2 = Bn

 $g: R^1 = H, R^2 = CH_0CH_0CO_0Bu^t$

oxazolidinone moiety followed by Cbz-protection of the free amino group was only viable for 4-isopropyl β -lactam 77 (Scheme 24).

Scheme 24

$$Xc \stackrel{H}{=} \stackrel{R^1}{=} R^2$$

$$SiMe_3$$

$$73$$

$$Cbz-HN \stackrel{H}{=} \stackrel{R^1}{=} R^2$$

$$O H$$

$$R^1 = H, R^2 = Pr^1$$

Reagents, conditions, and yields: i. (NH₄)₂Ce(NO₃)₆, MeOH, ~20 °C, 6 h, 80%; ii. 1) Li, NH₃, Bu¹OH, THF, 80%; 2) CbzCl, DMAP, 70%.

78: $R^1 = H$, $R^2 = CH_2CH_2Ph$ **79:** $R^1 = Me$, $R^2 = Me$

80: $R^1 = H$, $R^2 = CH_0CH_0Ph$, $R^3 = Bn$

81: $R^1 = H$, $R^2 = CH_2CH_2Ph$, $R^3 = CHMe_2$

82: $R^1 = Me$, $R^2 = Me$, $R^3 = Bn$

83: $R^1 = Me$, $R^2 = Me$, $R^3 = CHMe_2$

The approach to dipeptide products was guided by the observation that the introduction of the Boc group at the endocyclic N atom in β -lactams 76e and 76h proceeded chemoselectively to give the differently protected β -lactams 78 and 79, respectively (Scheme 25). Further, the presence of an electron-withdrawing group at this N atom also anticipated the expected enzymemimetic ring opening with α -amino acid esters. However, while the β -lactam 78 coupled efficiently with both (S)-phenylalanine methyl ester and (S)-valine methyl ester in DMF in the presence of NaN₃ promoter to give dipeptides 80 and 81 in good yields, the β -lactam 79 with a quaternary C(4) atom did not react with these α -amino acid esters, even

NaN₃ or KCN, DMF, ~20 °C, 70--89%; iii. H₂, Pd/C, EtOH, ~20 °C, 14 h, 90%.

when a twofold excess of NaN₃ was added. Nonetheless, the coupling reaction can be effected efficiently by replacing the additive NaN₃ with KCN. Under these conditions the dipeptide product 82 was produced within about 10 h in 89% isolated yield and dipeptide 83 in 85% yield after about 20 h of reaction.

Further examples are shown in Scheme 26. For example, acylation of \(\beta-lactam 84 with (S)-Cbz-leucine fluoride ((S)-Cbz-LeuF) in the presence of N-methylmorpoline (NMM) according to Carpino's procedure and subsequent introduction of N-Boc led to the β -lactam 85. This β-lactam dipeptide also coupled efficiently under the influence of either NaN3 or KCN with both (S)-phenylalanine methyl ester and (S)-valine methyl ester to give tripeptides 86 and 87 in 80 and 75% yields, respectively. When the β-lactam 85 was subjected to hydrogenolysis, the piperazinedione 88 was produced smoothly in 90% isolated yield. Clearly, these examples demonstrate that the approach developed can be readily extended to the synthesis of cyclic peptides, peptidomimetics, and biologically active substances incorporating α,β -diamino acids as key segments. 107

4. Macrocyclic peptides

There are several macrocyclic peptides containing non-natural α-amino acids, particularly α-amino-β-hydroxy acids. ¹⁰⁸ Most of these complex compounds have useful biological properties and often act as potent antibiotics. Two representative examples are vancomycin ¹⁰⁹ and lysobactin (89). ⁹⁹ The latter is a macrocyclic peptide lactone isolated from the fermentation of *Lysobacter* Sp.AICC53042 and shows a similar mode of antibacte-

Scheme 28

Reagents, conditions, and yields: *i.* 1) Cianuric fluoride, Py, CH_2Cl_2 , 6 h; 2) (S)- $H_2NC(CH_2OBn)CO_2Bn$, CH_2Cl_2 , NMM, 3 h, 95%; *ii.* 1) MeCN, H_2O , CH_2Cl_2 , 90%; 2) (Boc)₂O, MeCN, DMAP, 80%; *iii.* 1) NH₄OH, DMF, 90%; 2) BocGlyF, NMM, CH_2Cl_2 , 2 h, 0 °C, 80%.

rial action to that of vancomycin. Using the β -lactam approach to both α - and β -amino acids described in the earlier sections, the synthesis of the non-proteinogenic α -amino- β -hydroxyacid derived peptides 90 and 91 occurring in this antibiotic can be achieved with high diastereoselectivity and efficiency.

Synthesis of tripeptide **90** (Scheme 28) started from β -carboxy- β -lactam **92**, which is a form of β -hydroxy-aspartic acid possessing a β -carboxyl group and a α -amino moiety simultaneously protected. The dipeptide unit **93** was obtained in 95% overall yield after activation of the carboxy group with cyanuric fluoride and subsequent cou-

pling with O-benzyl-L-serine benzyl ester according to Carpino's procedure. Next, the β -lactam 93 was N-dearylated and converted into lactam 94 in good overall yield. The ring opening of 94 with 25% NH₄OH aq in DMF as solvent proceeded to give the expected product in 90% yield, which on N-Boc deprotection and further acylation with Boc-glycyl fluoride (Boc—GlyF) and NMM gave tripeptide 95 in 80% yield¹¹⁰ (for a conventional synthesis of β -hydroxyaspartyl dipeptides, see Ref. 111).

The strategy for the synthesis of tripetide 91 is shown in Schemes 29 and 30. The key step in this strategy was the anticipated stereocontrolled synthesis of β-lactams 97a and 97b incorporating the required structural subunits of the desired non-proteinogenic α-amino-β-hydroxy acids. These were prepared by cycloaddition of benzyloxyketene to imines 96a and 96b, respectively, followed by hydrogenolysis of the benzyloxy protective group. Subsequent exposure of both compounds to a solution of commercial bleach and a catalytic amount of TEMPO afforded NCAs 98a and 98b in 95 and 96% yields, respectively. The coupling reaction of 98a with (S)-leucine benzyl ester in methylene chloride as solvent proceeded cleanly to give dipeptide 99 in 90% yield after N-debenzylation of the resulting intermediate dipeptide product. Finally, as shown in Scheme 30, acylation of 99 with the NCA 98b carried out in DMF as solvent in the presence of NaN₃ gave the tripeptide 100 in 70% yield. 110

Scheme 29

OR

$$H_2N_{11}$$
 H_2N_{11}
 H

Reagents, conditions, and yields: *i.* 1) BnOCH₂COCl (2 equiv.), NEt₃, CH₂Cl₂, -78 °C→-20 °C, 20 h; 2) NH₄HCO₂, Pd/C, PrⁱOH, boiling, 1 h, 70–78%; *ii.* 1 M NaOCl, TEMPO (cat.), -20 °C, 95–96%; *iii.* 1) S-LeuOMe, CH₂Cl₂, -20 °C, 15 h, 95%; 2) H₂, Pd/C, EtOH, -20 °C, 15 h, 85%.

Reagents, conditions, and yield: i. DMF, NaN₃ (1 equiv.), -20 °C, 15 h, 70%.

The preceding synthesis of the non-proteinogenic β -hydroxy- α -amino acid peptides of lysobactin is just one example that demonstrates the potential utility of this approach to peptide segments found in other macrocyclic compounds. 8,108

5. 1,3-Polyols and amino polyols

In conjunction with the above studies we have also developed a β -lactam route to 2-amino-1,3-polyol chains using an iterative cycloaddition of alkoxyketenes to α -silyloxyaldehyde-derived imines as the key reaction. ¹¹² As Scheme 31 illustrates, the synthesis starts with the transformation of the imine 101 to β -lactam 102. Subsequent ring opening under mild acid conditions provided the α , γ -dialkoxy- β -amino ester 103 in almost quantitative yield, which was next converted into the isocyanide 104. Reduction of the latter proved to be difficult under Barton's conditions, but using Chatgilialoglu's reagent, tris(trimethylsilyl)silane, the reduction proceeded efficiently to give the 1,3-diol 105 in 80% yield. ¹¹²

The second cycle of the iterative process is shown in Scheme 32. The hydroxy derivative 106, obtained by reduction of ester 105 with lithium diisobutylaluminum hydride, was oxidized to the corresponding aldehyde and converted into the imine 107. Treatment of this imine with benzyloxyketene led to the β-lactam 108 in 75% yield as a single diastereomer. Subsequent N-dearylation and further ring opening provided the β-amino- α , γ, ε-polyol chain 109 in 50% overall yield. Deamination of 109 followed by repetition of the above sequence of reactions should generate more extended amino polyols. This synthesis nicely illustrates how a number of 1,3-diol subunits can be incorporated in a carbon chain by relying on the predictable stereochemistry provided by the cycloaddition reaction of alkoxyketenes with α-oxyaldehyde-derived imines.

DAM 101 BnO
$$\frac{H}{I}$$
 $\frac{H}{I}$ $\frac{$

Reagents, conditions, and yields: *i*. 1) BnOCH₂COCl, NEt₃, 80%; 2) CAN, MeCN, H₂O, 75%; *ii*. MeOH, ClSiMe₃, 0 °C \rightarrow 20 °C, 2 h, 98%; *iii*. 1) Ac₂O-HCO₂H, 85%; 2) (Cl₃CO)₂CO, NEt₃, CH₂Cl₂, 0 °C, 15 h, *iv*. (SiMe₃)₃SiH, AIBN, toluene, 80 °C, 30 min, 80%.

Scheme 32

Reagents, conditions, and yields: *i.* 1) (Cl₃CO)₂CO, DMSO, NEt₃, CH₂Cl₂, 80%; 2) 4-MeOC₆H₄NH₂, CH₂Cl₂, MgSO₄, 100%; *ii.* BnOCH₂COCl, NEt₃, 75%; *iii.* 1) CAN, MeCN, H₂O, 50%; 2) MeOH, ClSiMe₃, 0 °C → ~20 °C, 2 h, 100%.

6. β-Amino ketones and derived hydroxy compounds

Because of the general interest in aminopolyols and biologically active peptides, another approach to these attractive targets has also been delineated in our laboratory. The approach was based on the successful implementation of the N(1)—C(2) β -lactam cleavage by means of heteronucleophiles discussed in the earlier sections. On the one hand, we expected that ring opening of β -(1-alkoxyalkyl)-substituted β -lactams by carbon nucleophiles would provide β -aminoketones, which after diastereoselective reduction would provide a route to more complex aminopolyols and, on the other hand, the same reaction carried out on N-Boc- β -lactams with alkyl substituents at the β -C atom would generate precursors of peptide isosteres and/or renin inhibitors.

Our approach to aminopolyols is illustrated in Scheme 33. The most significant feature is that the reaction of aryl Grignard reagents at low temperature (-40 °C) with the N-Boc- β -lactam 110, obtained from 63, affords exclusively β -amino ketones 111 in 90-96%

yields. Overaddition could not be observed under these conditions even when an excess of the reagent was present in the reaction medium. At the same time, when the reaction was performed at room temperature instead of -40 °C, only tertiary alcohols were obtained. In contrast to these observations, reaction of lactam 110 with primary alkylmagnesium halides as well as with alkyllithiums at -40 °C invariably led to a mixture of the corresponding β-amino ketones 111 and β-amino carbinols 112, along with the starting β-lactam. Alternatively, the use of Gilman cuprates afforded good results in terms of chemoselectivity, although the yields were only moderate (R = Me, 50%: R = Buⁿ, 67%). As shown in Scheme 33, β-amino ketones carrying additional functionality, such as 113 and 114, could also be prepared by this method in good yields.

Another example that illustrates the utility of this procedure is shown in the synthesis of the α -amino- γ -hydroxy acid 120 present in the nucleoside antibiotic nikkomycin (121) as a structural segment 114 (for a recent review of nucleoside antibiotics, see Ref. 115). The

Reagents, conditions, and yields: i. RMgX or RLi, THF, -40 °C, 1 h; ii. ArMgX (1.3 equiv.), THF, -40 °C, 1 h, 90%, or R₂CuLi, Et₂O, 0 °C→-20 °C; iii. CH₂=C(OLi)OMe (2 equiv.), THF, -40 °C, or PhMe₂SiCH₂MgCl, THF, -20 °C; iv. RMgX, -78 °C→~20 °C.

Scheme 34

Me

$$H = H = CO_2Me$$
 $H = CO_2Me$
 $H = CO_2M$

Reagents, conditions, and yields: i. RMgBr, THF, -40 °C, 60-80%; ii. LiBH(Bus)3, THF, -78 °C, 1 h, 73%; iii. NDC, Py, C₆H₆, ~20 °C, 4 h.

method takes advantage of the fact that Grignard reagents react with rac-115 selectively at the lactam carbonyl group to afford exclusively products 116 and 117. At the same time, while methylmagnesium bromide gave a mixture of 116a and 117a along with the starting compound 115 in a ratio 70:10:20, respectively, aryl Grignard reagents only gave \(\beta\)-amino ketones 116b and 116c in 80 and 12% yields, respectively. L-Selectride reduction of 116b gave the amino lactone 118 and the hemiacetal 119 in a ratio of 62: 3, respectively. This hemiacetal was oxidized to the amino lactone by using the mixed nicotinic-chromic anhydride (NDC) reagent developed in our laboratory. 116 By that means, the overall yield of 118 was 73%. This result illustrates the viability of the approach for the synthesis of β-alkylα-amino-γ-hydroxy acids in optically pure forms starting from homochiral *cis*- β -alkyl- γ -alkoxycarbonyl β -lactams.

This approach to β-amino ketones has also been extended to the synthesis of alkyl-substituted 1,2,3-amino diols. 1,117 As shown in Scheme 35, when lactam 122 is treated with isobutylmagnesium chloride, β-amino ketone 123 forms along with the corresponding overaddition product (10%). The reduction of 123 under chelationcontrolled conditions led to the corresponding carbinol together with its epimer in a ratio 90: 10, respectively. Hydrogenolysis of the major isomer and further saturation of the phenyl ring afforded the 1,2,3-aminodiol 124, a constituent of the potent inhibitor of human renin A-725-17 (125), which is undergoing clinical evaluation as a therapeutic agent for the treatment of hypertension and congestive heart failure. 119,120

125

Reagents, conditions, and yields: i. MgCl , THF, -40 °C, 70%; ii. 1) n-Bu₄NF; 2) NaBH₄; 3) H₂ (3.5 atm), Rh—Al.

7. Pyrrolizidine alkaloids

In the course of our studies on the cycloaddition of ketenes with N-Boc- α -aminoaldehyde-derived imines we observed that the resulting cycloadducts easily underwent intramolecular rearrangement by attack of the exocyclic amino group on the β -lactam carbonyl to form γ -lactams. This approach to five-membered rings has been exploited to obtain necine bases bearing the 1,2-aminoalcohol functionality, which occurs, for instance, in some pyrrolizidine alkaloids such as lolidine (126), absouline (127), and related systems. 123

For example (Scheme 36), treatment of methoxy-ketene, generated from methoxyacetyl chloride and triethylamine, with the prolinal imine 128 afforded the β -methoxy- β -lactam 130 in 85% yield and as a single diastereomer. Likewise, reaction of benzyloxyketene with both imines 128 and 129 proceeded with virtually com-

Scheme 36

Reagents, conditions, and yields: i. R¹OCH $_2$ COCl, NEt $_3$, CH $_2$ Cl $_2$, -78 °C \rightarrow ~20 °C, 70-85 %; ii. 1) (NH $_4$) $_2$ Ce(NO $_3$) $_6$, MeCN, H $_2$ O; 2) ClSiMe $_3$, MeOH, boiling or F $_3$ CCO $_2$ H. CH $_2$ Cl $_2$, then 12 N HCl, EtOH, boiling, 70%; iii. BH $_3$, SMe $_2$, THF, boiling, 2 h, NaOAc, MeOH, 5 min, then I $_2$, CHCl $_3$; iiii. (Boc) $_2$ O, NEt $_3$, CH $_2$ Cl $_2$, boiling.

plete diastereoselectivity to give lactams 131 and 132 in 75% and 70% yields, respectively. The absolute configuration given for the adducts is based on a single crystal X-ray analysis of lactam 130 and the assumption of a

uniform reaction mechanism. The β -lactam 131, when subjected to N-dearylation and further treatment with chlorotrimethylsilane in methanol, gave the γ -lactam 133, which was isolated as the N-Boc derivative 134. The acylation of 133 with (+)-MTPA acid chloride and triethylamine to form a single diastereomer proved its optical purity. Treatment of compound 134 with trifluoroacetic acid followed by 12N HCl in refluxing ethanol gave the bicyclic compound 135, which then was converted into the pyrrolizidine framework 136.

8. β, γ -Dihydroxy and γ -amino- β -hydroxy acids

On the basis of the results achieved on opening of the nucleophilic β-lactam ring, particularly by means of α-amino acid esters, the analogous reaction on β-lactones derived from both α -oxyaldehydes and α -amino aldehydes was also explored in our laboratory. In fact, two modes of \beta-lactone ring opening have been described arising from either acyl-oxygen or alkyl-oxygen bond fission but results achieved to date concerning the first mode of cleavage have not been uniformly good, 124-127 We reasoned that β-lactones bearing electron-withdrawing groups at the α-position would be able to undergo nucleophilic attack by α -amino acid esters to form peptides that include segments of β-hydroxycarboxylic acids. In particular, starting from β-lactones derived from a-amino aldehydes it would be possible to obtain biologically active peptide mimics. 128

As Scheme 37 illustrates, the reaction of dichloroketene, generated from dichloroacetyl chloride and triethylamine, with the N-Boc- α -amino aldehyde 137 afforded the β -lactone 138 in 30% yield. Exposure of this

Scheme 37

Reagents, conditions, and yields: i. $Cl_2CHCOCl$, NEt_3 , -78 °C \rightarrow -20 °C, 30-35%; ii. H_2 , Pd/C, NEt_3 , EtOAc, 98%; iii. H_2 , Pd/C, NEt_3 , EtOH, 98%.

β-lactone product to hydrogenolysis in EtOH for 20 h gave the known ester $140,^{129}$ which is the key component of some HIV protease inhibitors, vide infra. At the same time, when the dehalogenation of lactone 138 was conducted under non-solvolytic conditions, the β-lactone ring was preserved and product 139 was obtained in almost quantitative yield. In a similar way (Scheme 38) reaction of dichloroketene, generated from trichloroacetyl chloride and Zn/Cu, with aminoketene 141 followed by intramolecular cyclization afforded the γ-lactam 143 in 35% yield over the two steps. On dehalogenation, this compound gave the known hydroxy derivative 144, ¹³¹ thus proving both the stereochemical course of the cycloaddition and its synthetic utility.

Scheme 38

Reagents, conditions, and yields: i. Cl_3CCOCl , Zn/Cu, 35%; ii. $ClSiMe_3$, MeOH, 6 h, \geq 90%; iii. H_2 , Pd/C, EtOAc, NEt_3 , 90%.

We also have found (Scheme 39) that lactone 138 efficiently coupled with amino acid esters to form, after dehalogenation, the corresponding acyl derivatives such as 145 and 146 in good yields. Though a somewhat lower yield was observed in the coupling reaction of 138 with the dipeptide (S)-Val-ValOMe to give 147, these results show the potential of the approach to synthesize HIV protease inhibitors. In contrast, and in agreement with our initial hypothesis, the β -lactone 139 was inert to ring opening by α -amino acid esters under the same conditions, in which the ring of 138 clearly did open.

In addition to these results we also have found (Scheme 40) that slow addition of trichloroacetyl chloride to a mixture of α -silyloxy aldehydes 148 and Zn/Cu in diethyl ether furnished the corresponding β -lactones in 60-85% yields and in each case as single diastereomers. The coupling reaction of both 149a and 149b with (S)-ValOMe also proceeded efficiently to give, after hydrogenolysis and desilylation, the corresponding

Reagents, conditions, and yields: i. 1) H_2N CO_2Me , CH_2CI_2 , ~ 20 °C, 24 h; 2) H_2 , Pd/C, NEt_3 , EtOAc, ~ 20 °C, 24 h;

ii. 1)
$$H_2N$$
 CONH CO $_2$ Me , ~20 °C, 24 h; 2) H_2 , Pd/C, NEt $_3$, EtOAC, ~20 °C

Scheme 40

Reagents, conditions, and yields: i. Cl_3CCOCl , Zn/Cu, Et_2O , 4 h, ~20 °C, 60–85%; ii. 1) (S)-ValOMe, CH_2Cl_2 , 24 h; 2) $H_2(1 \text{ atm})$, Pd/C, NEt_3 , EtOAc, 15 h, ~20 °C; 3) n-Bu₄NF, THF.

acyl derivatives 150 and 151. Although this work is still in its preliminary stages the approach developed seems promising for the incorporation of β -hydroxycarboxylic acids into short peptide fragments. ^{132*}

9. Conclusion

It is evident from the results presented in the above sections that appropriately substituted \(\beta\)-lactam frameworks constitute effective tools for the incorporation of a wide variety of both β - and α -amino acids into short peptide segments. Owing to the reliable enzymemimetic coupling reaction of β -lactams with α -amino acid esters under racemization-free conditions, the synthesis of many peptide segments and derived products may be now considered as a routine task, whenever the adequate β-lactams can be prepared. Therefore, the forthcoming development of novel procedures to procure homochiral azetidin-2-ones bearing any substitution pattern by singlestep and convergent approaches, would be highly desirable. In addition to these aspects, the discovery that α hydroxy-β-lactams can be directly transformed into simultaneously amino-protected and carboxyl-activated forms of α -amino acids from non- α -amino acid precursors renders the β-lactam-NCA method a very promising tool for the synthesis of complex peptide targets.

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^{*} C. Palomo, J. I. Miranda, and A. Linden, Unpublished results.

References

- 1. C. Palomo, I. Ganboa, and J. M. Aizpurua, in *Book of Abstracts IXth European Symposium on Organic Chemistry* (June 18-23, 1995, Warsaw, Poland), p. 42.
- 2. W. F. Degrado, Adv. Protein Chem., 1988, 39, 51.
- 3. C. Toniolo, Int. Peptide Protein Res., 1990, 35, 287.
- 4. A. Giannis and T. Kolter, Angew. Chem. Int. Ed. Engl., 1993, 32, 1244.
- R. M. J. Liskamp, Recueil Trav. Chim. Pays Bas, 1994, 113, 1.
- 6. J. Gante, Angew. Chem. Int. Ed. Engl., 1994, 33, 1699.
- 7. Amino Acids, Peptides and Proteins, Royal Society of Chemistry: London, 1994, 25.
- 8. R. M. Williams, Synthesis of Optically Active α-Amino Acids, Pergamon, Oxford, 1989.
- α-Amino Acid Synthesis, Ed. H. J. O'Donell, Tetrahedron Symposia-in-print, Tetrahedron, 1988, 44, 5253.
- 10. R. O. Duthaler, Tetrahedron, 1994, 50, 1539.
- P. D. Bailey, J. Clayson, and A. N. Boa, Contemp. Org. Synth. 1995, 2, 173.
- M. B. Smith, in Methods of Non α-Amino Acid Synthesis, Marcel Dekker, New York, 1995.
- 13. E. Juaristi, D. Quintana, and J. Escalante, Aldrichimica Acta, 1994, 27, 3.
- 14. D. C. Cole, Tetrahedron, 1994, 50, 9517.
- H. Heimgartner, Angew. Chem. Int. Ed. Engl., 1991, 30, 238
- D. Obrecht, V. Bohdal, J. Daly, C. Lehmann, P. Schonholzer, and K. Muller, Tetrahedron, 1995, 51, 10883.
- 17. L. S. Hegedus, Acc. Chem. Res., 1995, 28, 299.
- J. Podlech and D. Seebach, Angew. Chem. Int. Ed. Engl., 1995, 34, 417.
- 19. J. Podlech and D. Seebach, Liebigs Ann., 1995, 1217.
- 20. M. Hatam, D. Tehranfar, and J. Martens, Synthesis, 1994, 619.
- F. P. Cossio, C. Lopez, M. Oiarbide, C. Palomo,
 D. Aparicio, and G. Rubiales, Tetrahedron Lett., 1988,
 29, 3133.
- 22. H. R. Kricheldorf, α-Amino Acid N-Carboxy-Anhydrides and Related Heterocycles, Springer-Verlag, Berlin, 1987.
- 23. T. J. Blacklock, R. Hirschmann, and D. F. Veber, *The Peptides*, Academic Press: New York, 1987, 9, 39.
- C. Palomo, J. M. Aizpurua, C. Cuevas, R. Urchegui, and A. Linden, J. Org. Chem., in press.
- J. Backes, in Houben-Weil Methoden der Organischen Chemie; Eds. E. Muller and O. Bayer; Thieme, Stuttgart, 1991, Band E16B, p. 31.
- 26. G. I. Georg and V. T. Ravikumar, in *The Organic Chemistry of β-Lactams*; Ed. G.I.Georg, VCH., New York, 1992, p. 295.
- Ojima, I. Habus, M. Zhao, M. Zucco, Y. H. Park,
 C. M. Sun, and T. Brigand, Tetrahedron, 1992, 48, 6985.
- I. Ojima, S. A. Kuduk, J. C. Slater, R. H. Gimi, and C. M. Sun, *Tetrahedron*, 1996, 52, 209.
- A. K. Mukerjee and A. K. Singh, Tetrahedron, 1978, 34, 1731.
- M. S. Manhas, D. R. Wagle, J. Chang, and A. K. Bose, Heterocycles, 1988, 27, 1755.
- 31.1. Ojima, in *The Organic Chemistry of β-Lactams*, Ed. G. I. Georg, VCH, New York, 1992, p. 197.
- 32. I. Ojima, Acc. Chem. Res., 1995, 28, 383.
- 33. S. Kano, T. Ebata, and S. Shibuya, *Chem. Pharm. Bull.*, 1979, 27, 2450.

- M. Shiogaki, N. Ishida, and S. Sato, Bull. Chem. Soc. Jpn., 1989, 62, 3950.
- 35. J. G. Galluci, D. C. Ha, and D. J. Hart, *Tetrahedron*, 1989, 45, 1283.
- 36. J. E. Baldwin, R. H. Adlington, C. R. A. Godfrey, D. W. Gollins, M. L. Smith, and A. T. Russel, *Synlett*, 1993, 51.
- J. E. Baldwin, R. H. Adlington, C. R. A. Godfrey, D. W. Gollins, and J. G. Vaughan, J. Chem. Soc. Chem. Commun., 1993, 1434.
- D. M. Spero, S. Kapadia, and V. Farina, *Tetrahedron Lett.*, 1995, 36, 4543.
- I. Ojima, E. K. Ng, and C. M. Sun, Tetrahedron Lett., 1995, 36, 4547.
- 40. C. N. C. Drey, J. Lawbridge, and R. J. Ridge, J. Chem. Soc. Perkin Trans. 1, 1973, 2001.
- 41. I. Ojima, C. M. Sun, and Y. H. Park, J. Org. Chem., 1994, 59, 1249.
- 42. J. P. Greenstein and M. Winitz, Chemistry of the Amino Acids; John Wiley, New York, 1961, 2, p. 861.
- W. H. Daly and D. Poche, *Tetrahedron Lett.*, 1988, 29, 5859.
- 44. W. D. Fuller, M. P. Cohen, M. Shakankareh, and R. K. Blair, *J. Am. Chem. Soc.*, 1990, **112**, 7414.
- 45. J. Savrda and M. J. Wakselman, J. Chem. Soc. Chem. Commun., 1992, 812.
- R. Wilder and S. Mobashery, J. Org. Chem., 1992, 57, 2755.
- C. Schierlinger and K. Burger, Tetrahedron Lett., 1992, 33, 193.
- 48. Ch.-N. Hsiao and T. Kolasa, Tetrahedron Lett., 1992, 33, 269.
- 49. E. Frerort, J. Caste, J. Poncet, and P. Jouin, *Tetrahedron Lett.*, 1992, 33, 2815.
- O. Itoh, T. Honnami, A. Amiano, K. Muruta, Y. Koichi, and T. Sugita, J. Org. Chem., 1992, 57, 7334.
- 51. Ch.-B. Yue and F. Naider, J. Org. Chem., 1993, 58, 350.
- M. Hesse, in Ring Enlargement in Organic Chemistry, VCH: Weinheim, 1991.
- J. E. Baldwin, M. F. Chan, G. Gallacher, H. Otsuka,
 P. Monk, and K. Prout, Tetrahedron, 1984, 40, 4513.
- 54. P. J. Dunn, R. Haner, and H. Rapoport, J. Org. Chem., 1990, 55, 5017.
- 55. Y. Nakamura and C. Shin, Chem. Lett., 1992, 49.
- D. Seebach, A. Studer, E. Pfammatter, and H. Widner, Helv. Chim. Acta, 1994, 77, 2035.
- D. F. Rane, V. M.Girijavallabhan, A. K. Gavguly, R. E. Pike, A. K. Saksena, and A. T. McPhail, *Tetrahedron Lett.*, 1993, 34, 3201.
- 58. W. Hartwig and L. Mittendorf, Synthesis, 1991, 939.
- 59. J. E. Baldwin, R. M. Adlington, and D. J. Birch, J. Chem. Soc. Chem. Commun., 1985, 256.
- Y. Aoyagi, M. S. Chorgade, A. A. Padmapriya, H. Suguna, and S. M. Hecht, J. Org. Chem., 1990, 55, 6291.
- 61. 1. Ojima and Y. Pei, Tetrahedron Lett., 1990, 31, 977.
- 62. R. Badorrey, C. Cativiela, M. D. Diaz-de-Villegas, and J. A. Galvez, *Tetrahedron: Asymmetry*, 1995, **6**, 2787.
- R. Herranz, S. Vinuesa, J. Castro-Pichel, C. Perez, and M. T. Garcia Lopez, J. Chem. Soc. Perkin Trans. 1, 1992, 1825.
- 64. M. Wang and S. J. Gould, J. Org. Chem., 1993, 58, 5176.
- 65. Y. Umezawa, H. Morishima, S. Saito, T. Takita, H. Umezawa, S. Kobayashi, M. Otsuka, M. Narita, and M. Ohno, J. Am. Chem. Soc., 1980, 102, 6630.
- H. Arai, W. K. Hagmann, H. Suguna, and S. M. Hecht, J. Am. Chem. Soc., 1980, 102, 6631.

- 67. T. Takemoto, T. Nakajima, S. Arihara, and K. Koike, Yakugaku Zasshi, 1975, 95, 326.
- 68. J. F. Flippen and R. D. Gilardi, Acta Crystallogr. Sect. B, 1976, 32, 951.
- 69. J. M. Dewar and G. Shaw, J. Chem. Soc., 1962, 583.
- 70. I. Murakoshi, F. Ikegami, Y. Hinuma, and Y. Hanma, *Phytochemistry*, 1984, 23, 1905.
- I. H. Gilbert, D. C. Rees, A. K. Crockett, and R. C. F. Jones, *Tetrahedron*, 1995, 51, 6315.
- 72. R. C. F. Jones, A. I. Crockett, D. C. Rees, and I. H. Gilbert, *Tetrahedron Asymmetry*, 1994, 5, 1661.
- 73. I. Gilbert, D. C. Rees, and R. S. Richardson, *Tetrahedron Lett.*, 1991, 32, 2277.
- 74. R. C. F. Jones and G. J. Ward, *Tetrahedron Lett.*, 1988, 29, 3853.
- C. Palomo, F. P. Cossio, C. Cuevas, B. Lecea, A. Mielgo, P. Roman, A. Luque, and M. Martinez-Ripoll, J. Am. Chem. Soc., 1992, 114, 9360.
- C. Palomo, J. M. Aizpurua, F. Cabre, C. Cuevas, S. Munt, and J. M. Odriozola, *Tetrahedron Lett.*, 1994, 35, 2725.
- 77. C. Palomo, J. M. Aizpurua, F. Cabre, J. M. Garcia, and J. M. Odriozola, *Tetrahedron Lett.*, 1994, 35, 2721.
- E. Altmann, K.-H. Altmann, and M. Mutter, Angew. Chem. Int. Ed. Engl., 1988, 27, 858.
- D. Seebach, T. L. Sommerfeld, Q. Jiang, and L. M. Venanci, Helv. Chem. Acta, 1994, 77, 133.
- 80. T. Wohr and M. Mutter, Tetrahedron Lett., 1995, 36, 3847.
- 81. C. Palomo, J. M. Aizpurua, R. Urchegui, and J. M. Garcia, J. Chem. Soc. Chem. Commun., 1995, 2327.
- C. Palomo, J. M. Aizpurua, I. Ganboa, E. Maneiro, and B. Odriozola, J. Chem. Soc. Chem. Commun., 1994, 1505.
- D. R. Wagle, G. Garai, J. Chiang, M. G. Monteleone,
 B. E. Kurys, T. W. Strohmeyer, V. R. Hedge, M. S. Manhas, and A. K. Bose, J. Org. Chem., 1988, 29, 5065.
- 84. R. Annunziata, M. Benaglia, M. Cinquini, F. Cozzi, and F. Ponzini, J. Org. Chem., 1993, 58, 4746.
- M. Jayaraman, A. R. A. S. Deshmukh, and B. M. Bhawal, J. Org. Chem., 1994, 59, 932.
- C. Palomo, J. M. Aizpurua, I. Ganboa, B. Odriozola, R. Urchegui, and H. Gorls, J. Chem. Soc. Chem. Commun., in press.
- 87. The Chemistry of β-Lactams, Ed. M. I. Page; Blackie: London, 1992.
- 88. C. Palomo, J. M. Aizpurua, C. Cuevas, A. Mielgo, and R. Galarza, *Tetrahedron Lett.*, 1995, 36, 9027.
- R. A. Holton, Eur. Pat. Appl. EP400, 971, 1990; Chem. Abstr., 1990, 114, 164568q.
- Ojima, I. Habus, M. Zhao, M. Zucco, Y. H. Park,
 C. H. Sun, and T. Brigaud, Tetrahedron, 1992, 48, 6985.
- 91. V. Knott-Hunziker, S. Petturson, S. G. Waley, B. Jaurin, and T. Grundstrom, *Biochem. J.*, 1982, 207, 315.
- D. H. Rich, Peptidase Inhibitors in Comprehensive Medicinal Chemistry, Ed. P. G. Sammes; Pergamon, Oxford, 1990, 2, 391.
- 93. H. Umezawa, Small Molecular Immunomodifiers of Microbial Origin. Fundamental and Clinical Studies of Bestatin, Pergamon, Oxford, 1981.
- 94. H. Umezawa, Drug. Exptl. Cli. Res., 1984, 10, 519.
- 95. H. Blomgren and J. Wasserman, J. Canc. Lett., 1981,
- Matsumoto, Y. Kobayashi, Y. Takemoto, T. Kamijo, H. Harada, Y. Ito, and S. Terashima, *Tetrahedron*, 1992, 48, 1853.

- 97. C. Palomo, J. M. Aizpurua, and C. Cuevas, J. Chem. Soc. Chem. Commun., 1994, 1957.
- Ojima, C. M. Sun, and Y. H. Park, J. Org. Chem., 1994, 59, 1249.
- T. G. Hansson and J. Q. Kihlberg, J. Org. Chem., 1986,
 51, 4490.
- 100. J. O'Sullivan, J. E. McCullough, A. A. Tymiak, D. R. Kirsch, W. H. Trejo, and P. A. Principe, J. Antibiot., 1988, 41, 1740.
- 101. J. Kato, H. Hinoo, Y. Tervi, J. Kikuchi, and J. Shoji, J. Antibiot., 1988, 41, 719.
- 102. T. Kato, J. Antibiot., 1989, 42, C-2.
- 103. A. A. Tymiak, T. J. McCormick, and S. E. Unger, J. Org. Chem., 1989, 54, 1149.
- 104. C. Palomo, F. Cabre, and J. M. Ontoria, *Tetrahedron Lett.*, 1992, 33, 4819.
- 105. F. H. Van der Steen and G. Van Koten, *Tetrahedron*, 1991, 47, 7503.
- 106. C. Palomo, J. M. Aizpurua, M. Legido, R. Galarza, P. Deya, J. Dunogues, J. P. Picard, A. Ricci, and G. Seconi, Angew. Chem. Int. Ed. Engl., in press.
- C. Palomo, J. M. Aizpurua, R. Galarza, and A. Mielgo, J. Chem. Soc. Chem. Commun., 1996, 633.
- 108. A. Saeed and D. W. Young, Tetrahedron, 1992, 48, 2507.
- 109. A. V. R. Rao, M. K. Gurjar, K. L. Reddy, and A. S. Rao, Chem. Rev., 1995, 95, 2135.
- 110. C. Palomo, J. M. Aizpurua, I. Ganboa, B. Odriozola, M. Maneiro, J. I. Miranda, and R. Urchegui, J. Chem. Soc. Chem. Commun., 1996, 161.
- 111. Y. Liwschitz, A. Singerman, and S. Sokoloff, J. Chem. Soc. (C), 1968, 1843.
- 112. C. Palomo, J. M. Aizpurua, R. Urchegui, and J. M. Garcia, J. Org. Chem., 1993, 58, 1646.
- 113. C. Palomo, J. M. Aizpurua, J. M. Garcia, M. Iturburu, and J. M. Odriozola, *J. Org. Chem.*, 1994, 59, 5184.
- 114. A. G. M. Barret and S. A. Lebold, J. Org. Chem., 1990, 55, 5812 and references cited therein.
- 115. S. Knapp, Chem. Rev., 1995, 95, 1859.
- F. P. Cossio, M. C. Lopez, and C. Palomo, *Tetrahedron*, 1987, 43, 3963.
- 117. D. M. Spero, S. Kapadia, and V. Farina, *Tetrahedron Lett.*, 1995, 36, 4543.
- 118. I. Ojima, E. W. Ng, and C. M. Sun, Tetrahedron Lett., 1995, 36, 4547.
- 119. H. D. Kleinert, S. H. Rosenberg, V. Klinghofer, J. Barlow, K. Spina, J. Polakowski, P. Kovar, J. Cohen, and J. Denissen, Science, 1992, 257, 1940.
- 120. W. R. Baker and S. L. Condon, *J. Org. Chem.*, 1993, 58, 3277.
- 121. C. Palomo, F. P. Cossio, C. Cuevas, J. M. Odriozola, and J. M. Ontoria, *Tetrahedron Lett.*, 1992, 33, 4827.
- 122. C. Palomo, J. M. Aizpurua, C. Cuevas, P. Roman, A. Luque, and M. Martinez-Ripoll, Anales de Quimica Int. Ed., in press.
- 123. Naturally Occurring Pyrrolizidine Alkaloids, Ed. A. F. H. Rizk; CRC Press, Boston, 1991.
- 124. A. Griesbeck and D. Seebach, Helv. Chim. Acta, 1987, 70, 1325.
- 125. S. H. Rosenberg, S. A. Boyd, and R. A. Mantei, *Tetrahedron Lett.*, 1991, 32, 6507.
- 126. E. S. Ratemi and J. C. Vederas, *Tetrahedron Lett.*, 1994, 35, 7605
- 127. H. Shao, S. H. H. Wang, C.-W. Lee, G. Osapay, and M. Joodman, J. Org. Chem., 1995, 60, 2956.
- 128. R. M. Williams, in Biologically Active Peptides: Design,

- Synthesis and Utilization; Eds. W. V. Williams, and D. B. Weiner; Technomic: Lancaster, 1993, I, 187.
- 129. T. Nishi, M. Kitamura, T. Ohkuma, and R. Noyori,
- Tetrahedron Lett., 1988, 29, 6327.
 130. T. Yokomatsu, Y. Yuasa, and S. Shibuya, Heterocycles, 1992, 33, 1051.
- 131. G. B. Dreyer, B. W. Metcalf, T. A. Tomaszeck, T. J. Carr, A. C. Chandler, L. J. Hyland, S. A. Fakhoury,
- V. A. Magaard, M. L. Moore, J. E. Strickler, C. Debouck, and T. D. Merck, Proc. Natl. Acad. Sci. USA, 1989, 86, 9752.
- 132. C. Palomo, J. I. Miranda, C. Cuevas, and J. M. Odriozola, J. Chem. Soc. Chem. Commun., 1995, 1735.

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